

Syntheses and Anticholinesterase Activities of (3aS)-*N*¹,*N*⁸-Bisnorpheneserine, (3aS)-*N*¹,*N*⁸-Bisnorphysostigmine, Their Antipodal Isomers, and Other Potential Metabolites of Phenserine[†]

Qian-sheng Yu, Nigel H. Greig,* and Harold W. Holloway

Drug Design & Development, Laboratory of Cellular and Molecular Biology, Gerontology Research Center (4E02), National Institute on Aging, Intramural Research Program, National Institutes of Health, 5600 Nathan Shock Drive, Baltimore, Maryland 21224-6825

Arnold Brossi

School of Pharmacy, University of North Carolina at Chapel Hill, North Carolina 27599-7361

Received January 22, 1998

Hydrolysis of the carbamate side chains in phenserine [(-)1] and physostigmine [(-)2] yields the metabolite (-)-eseroline (3), and the red dye rubreserine (4) on air oxidation of the former compound. Both compounds lacked anticholinesterase activity in concentrations up to 30 mM, which would be unachievable in vivo. A second group of potential metabolites of 1 and 2 are the *N*¹,*N*⁸-bisnorcarbamates (-)9 and (-)10, prepared from (3aS)-*N*⁸-benzylnoresermethole (-)12 by the carbinolamine route. These entirely novel compounds proved to be highly potent inhibitors of acetylcholinesterase [(-)9] and of acetyl- and butyrylcholinesterase (AChE and BChE) [(-)10], respectively. To elucidate further the structure/anticholinesterase activity relationship of the described compounds, the antipodal isomers (3aR)-*N*¹,*N*⁸-bisnorcarbamates (+)9 and (+)10 were likewise synthesized from (3aR)-*N*⁸-benzylnoresermethole (+)12 and assessed. The compounds possessed moderate but less potent anticholinesterase activity, with the same selectivity as their 3aS enantiomers. Finally, the anticholinesterase activities of intermediates *N*¹,*N*⁸-bisnorbenzylcarbamates (-)18, (-) 19, (+)18, and (+)19, also novel compounds, were additionally measured. The 3aS enantiomers proved to be potent and selective inhibitors of BChE, particularly (-)19, whereas the antipodal isomers lacked activity.

Phenserine [(-)1], the phenyl carbamate analogue of physostigmine [(-)2], is a new and selective inhibitor of acetylcholinesterase (AChE) with minimal butyrylcholinesterase (BChE) action which is about to enter clinical trials for the treatment of Alzheimer's disease.^{1,2} The compound is sufficiently long acting for once daily administration and preferentially enters the brain, the site of Alzheimer's disease, reaching and maintaining a brain/plasma ratio of 10:1.^{1,3} As a consequence of these incorporated design features, the compound has an unusually wide therapeutic window, a highly favorable toxicological profile in dogs and rodents, and dramatic action in animal cognitive models.^{1,3} Additionally, it modulates the molecular events involved in the neuropathology of Alzheimer's disease, reducing the synthesis and secretion of β -amyloid precursor protein, the source of the Alzheimer neurotoxic peptide β -amyloid, both in in vitro and in vivo studies.^{3,4}

An attractive and pivotal point in the development of phenserine as a therapeutic is the assessment and synthesis of its potential metabolites and elucidation of their biological properties. Since hydrolysis of carbamyl esters and N-demethylation are well-established metabolic pathways,^{5,6} we thought it prudent to prepare the

key tricyclic compounds derived from phenserine [(-)1] and physostigmine [(-)2] by such action and evaluate their anticholinesterase activities (Figure 1).

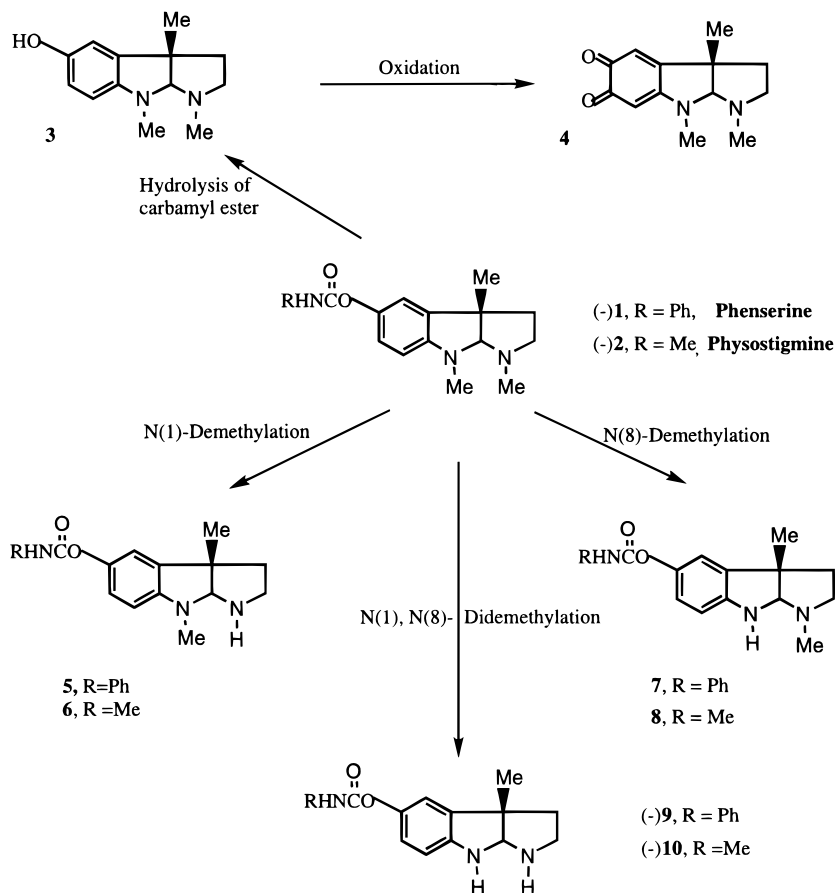
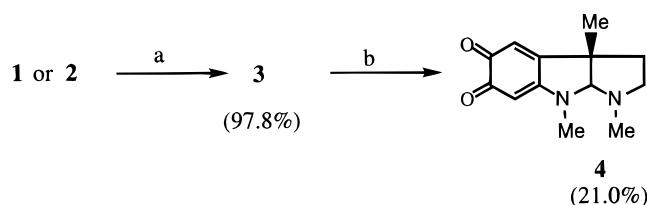
On the basis of our recent successful syntheses of the *N*¹-nor series⁷ and *N*⁸-nor series⁸ of both phenserine and physostigmine, we accomplished the novel syntheses of (3aS)-*N*¹,*N*⁸-bisnorpheneserine [(-)9] and (3aS)-*N*¹,*N*⁸-bisnorphysostigmine [(-)10]. As our earlier studies proved that the 3aS enantioselectivity is clearly maintained in the *N*⁸-nor series⁸ but not in the *N*¹-nor series,⁹ we additionally synthesized the (3aR)-*N*¹,*N*⁸-bisnor compounds (+)9 and (+)10 to assess their anticholinesterase activities, and report on them herein. Finally, no report on the synthesis, and chemical and biological evaluation of potential metabolites of phenserine and physostigmine would be complete without the inclusion of (-)-eseroline (3), the hydrolysis product of (-)1 and (-)2, and the red dye rubreserine (4), which is readily formed on air oxidation of 3. The evaluation of these compounds is likewise reported herein and was assessed against freshly obtained and prepared human AChE and BChE in vitro, alongside and compared to the activities of (-)- and (+)-phenserine (1) and physostigmine (2).

Results

Chemistry. Rubreserine (4) was obtained on air oxidation of eseroline (3),¹⁰ instead of by its isolation from a mixture of degradation products from physo-

* To whom correspondence should be addressed. Phone: 410-558-8278. Fax: 410-558-8323. E-mail: GreigN@vax.grc.nia.nih.gov.

[†] This paper is dedicated to Dr. Nelson J. Leonard, Faculty Associate in Chemistry, California Institute of Technology, Pasadena, CA, on the occasion of his 80th birthday.

**Figure 1.****Scheme 1^a**

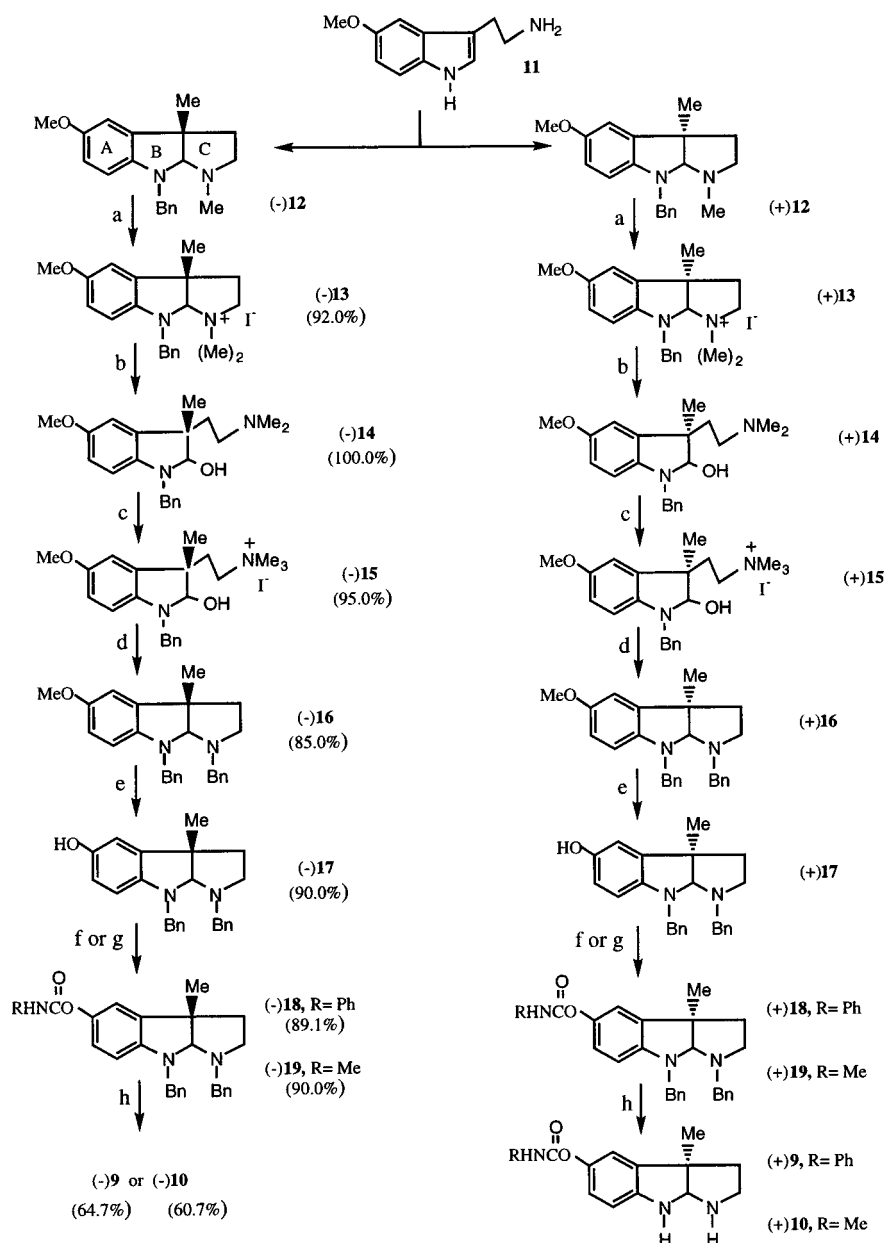
^a Reagents: (a) C₄H₉OH, Na; (b) H₂O, Na₂CO₃, O₂, CHCl₃.

stigmine [(-)2].^{11–14} Rubreserine (4) was initially isolated and identified by Auterhoff.¹¹ Later, Schöenberger et al. characterized it by X-ray diffraction analysis¹³ and, more recently, Cordell provided its ¹H and ¹³C NMR.¹⁴ The synthesis of compound 4 is described in this paper (Scheme 1).

The major task in this project, which had not previously been accomplished, was the synthesis of *N*¹,*N*⁸-bisnorpheneserine [(-)9] and its methylcarbamate analogue (-)10, both obtainable from (-)1 and (-)2 by *N*-demethylation. We suspected that (3*a*,*S*)-*N*¹,*N*⁸-bisnoresermethole (not shown), obtained from 5-methoxytryptamine (11),¹⁵ was not a good substrate to complete our task as the corresponding phenol would be exquisitely sensitive to air oxidation and hence difficult to handle. Furthermore, the lack of selectivity of O–H over N–H for reaction with isocyanate would have resulted in undesired products and hence make the synthesis far more complex than necessary. We have therefore developed an alternative route to prepare the compounds of the *N*¹,*N*⁸-bisnor series, and this is shown in Scheme 2.

This route uses the known (-)-*N*⁸-benzylnoresermethole [(-)12], which was made from 11 in six steps including its optical resolution, as has been reported.⁸ For introducing a benzyl group into the *N*¹-position of precious optically pure compound (-)12, we adopted, step by step, a procedure reported in the literature.¹⁶ Starting with (-)12, it was quaternized with CH₃I, ring-C-opened under basic conditions, and again quaternized with CH₃I, leading via (-)13 to (-)14 and (-)15. Reaction of (-)15 with BnNH₂ directly provided *N*¹,*N*⁸-bisbenzylnoresermethole [(-)16] in a total of four steps with a final yield of 74%. Demethylation of (-)16 gave the phenol [(-)17] which, when reacted with either phenyl isocyanate or methyl isocyanate, afforded (-)-*N*¹,*N*⁸-bisbenzylnorpheneserine [(-)18] and (-)-*N*¹,*N*⁸-bisbenzylnorphysostigmine [(-)19], respectively.

The catalytic debenzoylation of (-)18 and (-)19 to give the desired bisnor compound (-)9 and (-)10, respectively, was accomplished over Pd(OH)₂/C using *i*-PrOH as a solvent. The (3*a*,*R*)-*N*¹,*N*⁸-bisnorpheneserine [(+)9] and (3*a*,*R*)-*N*¹,*N*⁸-bisnorphysostigmine [(+)10] were produced from (3*a*,*R*)-*N*⁸-benzylnoresermethole [(+)12] in the same manner, as shown in Scheme 2. This debenzoylation proved unexpectedly difficult and is summarized in Scheme 3. First, we tried the conditions used in our previous preparation of the *N*⁸-nor series.⁸ The debenzoylation of (-)18 was initially attempted in an acidic medium of CF₃COOH/CH₃OH/H₂O over the catalyst Pd(OH)₂/C. After 1.5 h of hydrogenation at atmospheric pressure, only one product, the ring-open β-2'-aminoethyl indole, compound 20, could be isolated from the reaction mixture. This suggests that ring C

Scheme 2^a

^a Reagents: (a) CH₃I, ether; (b) EtOH, NaOH (50%); (c) CH₃I, Et₂O; (d) BnNH₂; (e) BBr₃, CH₂Cl₂; (f) phenyl isocyanate, Et₂O, Na; (g) methyl isocyanate, Et₂O, Na; (h) Pd(OH)₂/C, *i*-PrOH.

opening of the *N*¹,*N*⁸-bisnor compounds formed during hydrogenation occurs much more readily than does that of the *N*⁸-nor compounds.

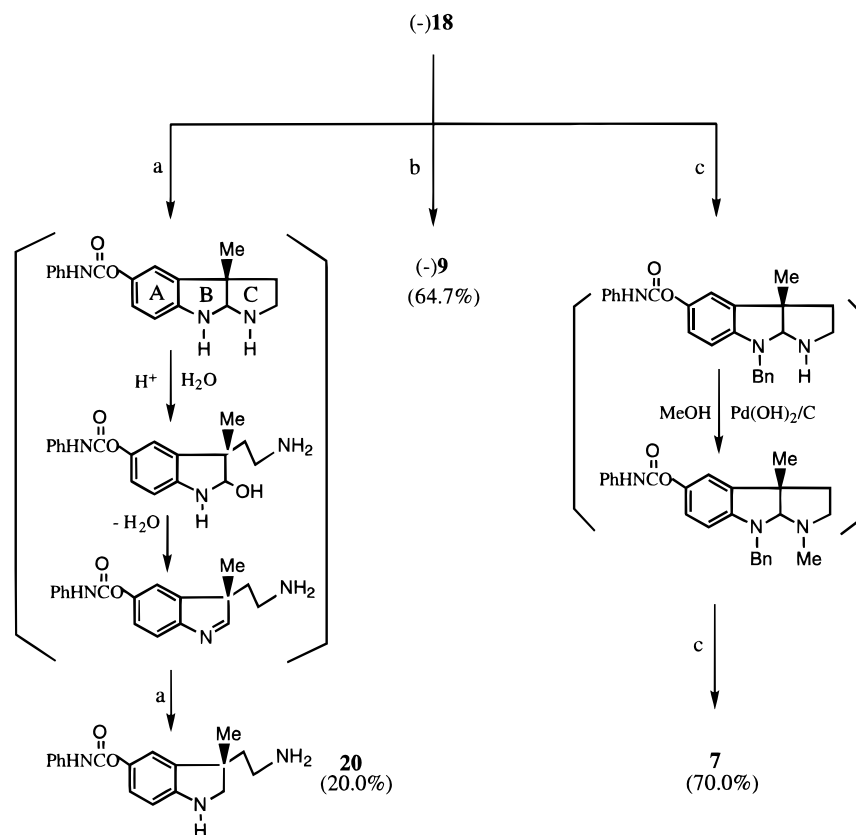
Extensive experimentation culminated in our use of a neutral solvent, initially CH₃OH, which, after 60 h of hydrogenation, provided (-)-*N*⁸-norphenserine 7, the only product that we were able to isolate from the reaction mixture. This suggests that during the long period of catalytic hydrogenation the initially formed *N*¹-nor compound was *N*-methylated, leaving the *N*⁸-benzyl group attached to the aniline nitrogen before finally giving the *N*⁸-nor compound 7. This result is in accord with studies by Baiker and Richarz¹⁷ and by He and Bossi.¹⁸ They described the alkylation of secondary amines on catalytic hydrogenation using a primary alcohol as the solvent at high temperature and proposed that the key intermediate of this reaction is the aldehyde formed from dehydrogenation of the primary

alcohol. We tried numerous solvents other than CH₃OH to avoid *N*¹-methylation, but these proved unsuccessful. Finally, we utilized *i*-PrOH or *t*-BuOH as the reaction solvents, and both provided positive results (Scheme 3).

Biological Evaluation. Table 1 illustrates the anticholinesterase activity of compounds 3, 4, (-)- and (+)9, 10, 18, and 19 against human AChE and BChE, compared to those of (-)- and (+)-phenserine (1) and physostigmine (2).

In accord with previous reports, neither (-)-eseroline (30) nor (-)-rubreserine (4) possessed anticholinesterase action. In contrast, the *N*¹,*N*⁸-bisnorcarbamates (-)9 and (-)10 were highly potent in this regard.

(-)-*N*¹,*N*⁸-Bisnorphenserine [(-)9] demonstrated potent and selective AChE inhibitory action with an IC₅₀ value similar to that of (-)-phenserine [(-)1]. Like (-)-phenserine, the compound lacked BChE inhibitory potency, and had a selectivity of AChE action of some

Scheme 3^a

^a Reagents: (a) Pd(OH)₂/C, H₂O, CH₃OH, TFA, 1.5 h; (b) Pd(OH)₂/C, *t*-PrOH, 60 h; (c) Pd(OH)₂/C, MeOH, 60 h.

Table 1. 50% Inhibitory Concentration (IC₅₀, nM) + SEM^a of Compounds versus Human Erythrocyte AChE and Plasma BChE

compound	IC ₅₀ AChE (nM)	IC ₅₀ BChE (nM)
(-) 1 , (-)-phenenserine	24.0 ± 6.0 ^a	1300 ± 85 ^a
(+) 1 , (+)-phenenserine	3500 ± 55	23500 ± 300
(-) 2 , (-)-physostigmine	27.9 ± 2.4	16.0 ± 2.9
(+) 2 , (+)-physostigmine	9890 ± 6	2490 ± 290
3 , (-)-eseroline	NDA ^b	NDA
4 , (-)-rubreserine	NDA	NDA
(-) 9 , (-)- <i>N</i> ¹ , <i>N</i> ⁸ -bisnorphenenserine	22.1 ± 2.3	897 ± 104
(+) 9 , (+)- <i>N</i> ¹ , <i>N</i> ⁸ -bisnorphenenserine	231 ± 23	952 ± 107
(-) 10 , (-)- <i>N</i> ¹ , <i>N</i> ⁸ -bisnorphysostigmine	10.9 ± 1.2	2.4 ± 0.8
(+) 10 , (+)- <i>N</i> ¹ , <i>N</i> ⁸ -bisnorphysostigmine	1490 ± 120	237 ± 55
(-) 18 , (-)- <i>N</i> ¹ , <i>N</i> ⁸ -bisbenzylnorphenenserine	41130 ± 11760	513 ± 311
(+) 18 , (+)- <i>N</i> ¹ , <i>N</i> ⁸ -bisbenzylnorphenenserine	NDA	NDA
(-) 19 , (-)- <i>N</i> ¹ , <i>N</i> ⁸ -bisbenzylnorphysostigmine	1090 ± 123	8.1 ± 2.1
(+) 19 , (+)- <i>N</i> ¹ , <i>N</i> ⁸ -bisbenzylnorphysostigmine	3560 ± 400	776 ± 312
5 , (-)- <i>N</i> ¹ -norphenenserine	13.8 ± 0.7	612 ± 1
6 , (-)- <i>N</i> ¹ -norphysostigmine	21.0 ± 1.0	2.0 ± 1.0
7 , (-)- <i>N</i> ⁸ -norphenenserine	40.8 ± 3.4	516 ± 601
8 , (-)- <i>N</i> ⁸ -norphysostigmine	56.7 ± 4.4	6.8 ± 2.2
21 , (-)- <i>N</i> ⁸ -benzylnorphysostigmine	330 ± 100	10.4 ± 4.0
22 , (-)- <i>N</i> ¹ -benzylnorphysostigmine	161 ± 41	21.8 ± 10.9

^a Standard error of the mean. ^b NDA: no detectable activity (at 3 × 10⁻⁵ M).

40-fold. In stark contrast to (+)-phenenserine [(+)**1**] which lacks anticholinesterase activity, (+)-*N*¹,*N*⁸-bisnorphenenserine [(+)**9**] possessed a low but nevertheless significant potency of AChE inhibition. The IC₅₀ value of this was 10-fold lower than that of (-)**9** for AChE, whereas the BChE inhibitory action of (+) and (-)**9** were similar and low.

(-)-*N*¹,*N*⁸-Bisnorphysostigmine [(-)**10**] possessed a high potency of inhibitory action against both AChE and BChE, with activity some 3- and 6-fold greater than that of (-)-physostigmine [(-)**2**] against the two enzyme subtypes, respectively. In contrast to (+)-physostigmine

[(+)**2**] which entirely lacked anticholinesterase action, (+)-*N*¹,*N*⁸-bisnorphenenserine [(+)**9**] proved inactive against AChE but possessed low BChE inhibitory action, quantitatively similar to that of (-)**9**.

The AChE inhibitory activity of the intermediates (-)-*N*¹,*N*⁸-bisbenzylnorphenenserine [(-)**18**] and (-)-*N*¹,*N*⁸-bisbenzylnorphysostigmine [(-)**19**] were dramatically reduced compared to (-)**9** and (-)**10**, respectively. However, their respective BChE inhibitory action was maintained, with (-)**19** being particularly potent. In contrast, the (+)-enantiomers were dramatically less active, with (+)-*N*¹,*N*⁸-bisbenzylnorphenenserine [(+)**18**]

entirely lacking demonstrable activity in concentrations up to 30 mM.

Discussion

We report, herein, the first synthesis and initial pharmacological evaluation of both natural and unnatural N^1, N^8 -bisnorpheneserine (**9**) and of N^1, N^8 -bisnorphysostigmine (**10**). Their structures and absolute configurations are secured with the chemical reactions described. Not only were the 3aS enantiomers of these novel compounds highly potent as anticholinesterases, but additionally they are potential metabolites of pheneserine [(-)**9**] and physostigmine [(-)**10**].

The classical anticholinesterase physostigmine [(-)**2**] has been utilized widely in clinical studies over the last several decades, and it as well as the quaternary carbamates such as neostigmine and pyridostigmine have demonstrated utility and acceptability in treating a variety of disorders, primarily involving an awry cholinergic system. These include paralytic ileus and atony of the bladder, glaucoma, myasthenia gravis, termination of the actions of competitive neuromuscular blocking agents, and overcoming intoxication with atropine, tricyclic antidepressants, or phenothiazines. Physostigmine is available for injection (antilirium), as an ointment, and, more recently, for experimental use in Alzheimer's disease as a slow-release formulation (synapton) that is in phase III clinical trials. As a first-generation drug, its major drawbacks in clinical use are its low therapeutic window and short duration of unselective action on acetyl- and butyrylcholinesterase, with pharmacokinetic and pharmacodynamic half-lives of 30–90 min, respectively.¹

In contrast, pheneserine [(-)**1**] and analogues overcome these disadvantages and possess a long duration of action (half-life 8–10 h) and an unusually wide therapeutic window which results in dramatic cognitive effects.^{1,3} Additionally, unlike physostigmine, pheneserine possesses the rare characteristic of modulating the processing of β -amyloid precursor protein to limit formation of the Alzheimer toxic peptide, β -amyloid.^{3,4} The agent therefore represents a promising drug candidate for the treatment of Alzheimer's disease as it has actions to potentially improve cognition and slow the disease course, through its interaction with the molecular events involved in the disease process, and its true value is about to be assessed in forthcoming clinical trials.

(-)- N^1, N^8 -Bisnorpheneserine [(-)**9**] and (-)- N^1, N^8 -bisnorphysostigmine [(-)**10**] both possessed potent anticholinesterase action and hence, if produced in vivo as a consequence of metabolism by N-demethylation, likely would potentiate or extend the action of (-)**1** or (-)**2**, respectively. Indeed, whereas (-)**9** is equipotent to (-)**1** versus AChE and BChE, with a slightly more modest AChE selectivity of 41-fold compared to 54-fold for (-)-pheneserine [(-)**1**], (-)- N^1, N^8 -bisnorphysostigmine [(-)**10**] proved to be more potent than (-)-physostigmine [(-)**2**]. Indeed, it was 6-fold more active in inhibiting BChE and 3-fold more potent against AChE, providing it a selectivity for BChE activity of some 5-fold, versus 2-fold for (-)-physostigmine [(-)**2**]. Thus, similar to (-)- N^1 and (-)- N^8 analogues, also potential metabolites of (-)**1** and (-)**2**, N-demethylation at both the N^1 and N^8 positions, separately and simul-

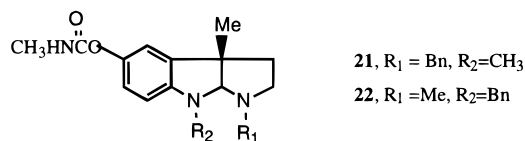
Table 2. Relative Selectivity of Compounds versus Human Erythrocyte AChE and Plasma BChE

compound	AChE selectivity	BChE selectivity
(-) 1 , (-)-pheneserine	54-fold	
(-) 2 , (-)-physostigmine		2-fold
5 , (-)- N^1 -norpheneserine	44-fold	
6 , (-)- N^1 -norphysostigmine		11-fold
21 , (-)- N^1 -benzylnorphysostigmine		32-fold
7 , (-)- N^8 -norpheneserine	13-fold	
8 , (-)- N^8 -norphysostigmine		8-fold
22 , (-)- N^8 -benzylnorphysostigmine		7-fold
(-) 9 , (-)- N^1, N^8 -bisnorpheneserine	41-fold	
(-) 10 , (-)- N^1, N^8 -bisnorphysostigmine		5-fold
(-) 19 , (-)- N^1, N^8 -bisbenzylnorphysostigmine		134-fold

taneously, results in highly potent anticholinesterases with a modest altered selectivity away from AChE (Table 2).

We have previously demonstrated that replacement of the methyl carbamate moiety of (-)-physostigmine [(-)**2**] with a substituted or unsubstituted phenylcarbamate,^{1,2,3,19} or with a longer alkyl carbamate, such as with a butyl, heptyl, or octyl carbamate, incrementally extends the duration of potent anticholinesterase action of the resulting compound.^{1,20,21} However, their longer duration of action likely results as a consequence of their higher rates of carbamylation of the cholinesterase enzyme and lower rates of decarbamylation,^{1,22,23} compared to (-)-physostigmine [(-)**2**], rather than as a consequence of their in vivo disappearance rates.¹ For example, as discussed, (-)-physostigmine [(-)**2**] has a duration of action of some 30–90 min and an in vivo disappearance half-life of 15 min.²⁴ The heptyl carbamate (Eptylstigmine, Mediolanum, Italy) in phase II clinical trials for the treatment of Alzheimer's disease, has a duration of action of in excess of 8 h and an in vivo disappearance half-life of 5 h.^{20,25} In contrast, (-)-pheneserine [(-)**1**], as discussed, likewise has a duration of cholinesterase inhibition of in excess of 8 h, but an in vivo disappearance half-life of only 12 min.^{1,3} It is therefore probable that in the event that (-)- N^1, N^8 -bisnorpheneserine [(-)**9**] and (-)- N^1, N^8 -bisnorphysostigmine [(-)**10**] are generated as metabolic products of (-)-pheneserine [(-)**1**] and (-)-physostigmine [(-)**2**] administration in clinical studies, such active metabolites would have a greater affect to potentiate the action of (-)**2** as its rate of enzyme decarbamylation is far more rapid than for (-)**1**. Although (-)-eseroline (**3**) has been reported to be pharmacologically active in antinociceptive assays,¹³ we reconfirm the lack of anticholinesterase activity of this known metabolite of (-)-physostigmine [(-)**2**] and of (-)-rubreserine (**4**), easily obtained from **3** on air oxidation and synthesized via a novel route.

We have previously demonstrated that inhibition of both AChE and BChE by (-)**1**, (-)**2**, and other N^1 -methyl-substituted carbamates is highly enantioselective, resting entirely on the 3aS enantiomers (Table 1).^{1,9} This, however, does not hold for the N^1 -nor series⁹ or for ring C heterocongeners,^{1,2,25,26} but does hold for the N^8 -nor series.⁸ Similar to the N^1 -nor series,⁹ (+)- $N(1,8)$ -bisnorpheneserine [(+)**9**] and (+)- $N(1,8)$ -bisnorphysostigmine [(+)**10**] retained anticholinesterase action, which was some 10-fold reduced compared to their 3aS enantiomers. Nevertheless, their activity is of the same magnitude as several anticholinesterases presently in

**Figure 2.**

clinical use: tacrine (Cognex, Warner-Lambert, MI) IC₅₀ AChE 190 nM, BChE 47 nM, and galanthamine (Johnson & Johnson, NJ) IC₅₀ AChE 800 nM, BChE 7300 nM.

We predicted from our previous studies involving modification of the N¹- and N⁸-positions of natural physostigmine [(−)**2**]^{1,2,8,27} that the intermediate (−)-N(1,8)-bisbenzylmorphysostigmine [(−)**19**] would possess anticholinesterase action with a selectivity for BChE, as **21** and **22** (Figure 2) proved to be BChE inhibitors equipotent to (−)**2** but with a 32-fold and 7-fold selectivity (Tables 1 and 2). We were, however, surprised by the remarkable potency and 134-fold BChE selectivity of (−)**19**. Our previous studies have demonstrated that the incorporation of a lipophilic group, such as a phenylethyl group,²⁸ in the N¹-position imparts a dramatic selectivity that favors BChE inhibitory action. The present studies clearly indicate that BChE inhibitory potency and selectivity can be increased by simultaneous lipophilic substitution in the N¹- and N⁸-positions; comparing the activity of (−)**19** to **21** and **22**. Additionally, they further suggest that BChE selectivity and, likely, duration of inhibition could be yet further increased by changing the methyl carbamate of (−)**19** to a 4'-substituted phenyl carbamate. We have previously demonstrated that such carbamates provide an extended duration of action, similar to that of (−)**1**, and a BChE selectivity.^{1,2,3,27}

In contrast, (−)-N(1,8)-bisbenzylmorphenserine [(−)**18**], which incorporates components to both favor BChE selectivity, in its lipophilic N(1,8)-bisbenzyl groups, and favor AChE selectivity, in its unsubstituted phenyl carbamate, possessed a minimal potency compared with (−)**1** and (−)**19** against both AChE and BChE. Likewise, the enantiomers (+)**18** and (+)**19** dramatically lacked activity, clearly indicating that whereas reduced steric factors associated with N¹-methyl substitution in the nor compounds,⁹ physovenols,²⁵ and thiaphysovenols²⁶ allow potent activity in the antipodal isomers, bulky substitutions do not.

A significant body of work has been published that describes the structures of AChE and BChE and their catalysis of acetylcholine (ACh) and related substrates.^{29,30,31} Both proteins have been fully sequenced, and although they are products of different genes on different chromosomes they share a remarkable degree of amino acid homology, particularly around the site involved in ACh hydrolysis.³² As we have described previously,⁸ it is clearly possible to design and synthesize analogues with specific substitutions that exploit critical differences around the active catalytic site that differentiate AChE from BChE to provide cholinesterase subtype inhibitory action for the resulting compounds.

In summary, we have synthesized and characterized the N(1,8)-bisnor analogues of both natural, (−)-**3aS**, and unnatural, (+)-**3aR**, physostigmine and phenserine. The **3aS** enantiomers, potential metabolites of (−)**1** and (−)**2**, proved to be highly active anticholinesterases, as did the intermediate (−)-N(1,8)-bisbenzylmorphysostig-

mine [(−)**19**], which demonstrated a surprising potency and selectivity for BChE. Whereas long-acting and nontoxic selective AChE inhibitors hold potential to augment memory processing and reduce β-amyloid synthesis in the treatment of Alzheimer's disease and age-associated memory deficits,^{24,33} recent studies have implicated BChE in the seeding and formation of β-amyloid plaques and its toxicity in Alzheimer's disease,^{34,35} providing a potential clinical use for interesting selective nontoxic BChE inhibitors.

Experimental Section

Chemistry. Melting points (uncorrected) were measured with a Fisher-Johns apparatus; ¹H NMR were recorded on a Bruker (Belleveca, MA) AC-300 spectrometer; MS (*m/z*) were recorded on a Hewlett-Packard 5890 GC-MS (EI) and on a Finnigan-1015D mass spectrometer. Optical rotations were measured by JASCO, model DIP-370 (Japan, Spectroscopic Co., Ltd.); elemental analyses were performed by Atlantic Microlab, Inc. Unless otherwise indicated, all separations were carried out using flash column chromatography (silica gel 60, 230–400 mesh) with the described solvents. All reactions involving nonaqueous solutions were performed under an inert atmosphere.

Rubreserine (4). The fumarate salt of eserine (**3**)¹⁰ (300 mg, 0.90 mmol) was dissolved in H₂O (10 mL). The solution was mixed with sodium carbonate (95 mg, 0.90 mmol) and CHCl₃ (20 mL). The reaction mixture was stirred under constant aeration, by filtering air flow through a small pipet for 20 h, and CHCl₃ was occasionally added to maintain a constant volume of CHCl₃. The separated CHCl₃ solution was washed with brine and dried over Na₂SO₄. After evaporation of solvent, the residue was chromatographed (CH₂Cl₂/MeOH = 20/1) and the visual red fraction was collected. Evaporation of solvent gave a residue which was crystallized from ether to give **4** (44 mg, 21.0%) as red crystals: mp 150–151 °C (lit.⁹ mp 152 °C; lit.¹² mp 98–99 °C); [α]_D²⁰ −256.3° (*c* = 0.2, CHCl₃); the ¹H NMR is identical with that reported in the literature (ref 14).

Methiodide of (−)-(3aS)-8-Benzyl-1,3a-dimethyl-1,2,3,3a,8,8a-hexahydro-5-methoxypyrrolo[2,3-*b*]indole [(−)13**].** Compound [(−)**12**]⁸ (180 mg, 0.58 mmol) was dissolved in ether (5 mL), and then CH₃I (0.5 mL) was added. The methiodide salt gradually formed and precipitated from the solution. The mixture was left overnight and filtered to give crystalline (−)**13** (240 mg, 92.0%): mp 140 °C; [α]_D²⁰ −109.0° (*c* = 0.5, EtOH). Anal. (C₁₉H₂₁N₃O₂) C, H, N.

(−)-(3s)-1-Benzyl-3-methyl-3-[2'-(dimethylamino)ethyl]-5-methoxy-2,3-dihydro-1H-indol-2-ol [(−)14**].** To a solution of (−)**13** (240 mg, 0.53 mmol) in H₂O (5 mL) and EtOH (1 mL) were added 50% NaOH (0.5 mL) and ether (5 mL). After the mixture was stirred at room temperature for 5 h, the ether layer was separated and the aqueous layer was washed with ether (2 × 5 mL). The combined ether layer then was washed with brine, dried over Na₂SO₄, and evaporated to give (−)**14** (182 mg, 100%) as a gum: [α]_D²⁰ −21.9° (*c* = 0.5, CHCl₃); ¹H NMR (CDCl₃) δ 7.40–7.15 (m, 5 H, Ph-H), 6.50 (m, 2H, C4-H, C6-H), 6.25 (d, *J* = 8.0 Hz, 1H, C7-H), 4.60 (s, 1H, C2-H), 4.45 and 4.15 (AB, *J* = 16.6 Hz, 2H, PhCH₂N), 3.60 (s, 3H, OCH₃), 2.15 (s, 6H, N-Me₂), 2.00–2.10 (m, 2H, CH₂N), 1.90–1.40 (m, 2H, C3-CH₂), 1.25 (s, 3H, C3-CH₃); EI-MS *m/z* (relative intensity) 322 (M⁺ − H₂O, 51), 307 (64), 264 (4.5), 91 (100). Anal. (C₃₃H₃₃N₃O₃·0.3H₂O) C, H, N.

Methiodide of (−)-(3s)-1-Benzyl-3-methyl-3-[2'-(dimethylamino)ethyl]-5-methoxy-2,3-dihydro-1H-indol-2-ol [(−)15**].** Compound (−)**14** (180 mg, 0.53 mmol) was dissolved in ether (5 mL), and then CH₃I (0.5 mL) was added. The methiodide salt gradually formed and precipitated from solution. The mixture was left overnight and filtered to obtain crystalline (−)**15** (243 mg, 95.0%): mp 172–173 °C; [α]_D²⁰ −391.0° (*c* = 0.2, EtOH). Anal. (C₁₉H₂₁N₃O₂·0.2H₂O) C, H, N.

(-)-(3a*S*)-1,8-Dibenzyl-3a-methyl-1,2,3,3a,8,8a-hexahydro-5-methoxypyrrolo[2,3-*b*]indole [(-)16]. Compound (-)15 (230 mg, 0.48 mmol) was dissolved in CH₃CN (5 mL), and benzylamine (103 mg, 0.96 mmol) was added. The reaction mixture was stirred and refluxed for 6 h. Evaporation of solvent gave a residue which was partitioned between H₂O and Et₂O. The ether layer was evaporated and chromatographed (CH₂Cl₂/MeOH = 20/1) to give (-)16 (157 mg, 85.0%) as a gum: [α]_D²⁰ -72.3° (*c* = 0.4, CHCl₃); ¹H NMR (CDCl₃) δ 7.30–7.20 (m, 10 H, Ar-H), 6.65 (d, *J* = 2.5 Hz, 1H, C4-H), 6.55 (d, *J* = 8.5 Hz, 1H, C6-H), 6.25 (d, 1H, *J* = 8.5 Hz, C7-H), 4.45 (s, 1H, C8a-H), 4.40 and 4.20 (AB, *J* = 16.6 Hz, 2H, PhCH₂-N8), 3.80 (m, 2H, PhCH₂-N1), 3.60 (s, 3H, OCH₃), 3.70 (s, 3H, CH₃O), 2.75 (t, *J* = 7 Hz, 2H, C2-H₂), 1.95 (t, *J* = 7 Hz, 2H, C3-H₂), 1.40 (s, 3H, C3a-CH₃); EI-MS *m/z* (relative intensity) 294 (MH⁺ - benzyl, 48), 279 (8.9), 188 (23), 173 (58), 91 (100). Anal. (C₂₆H₂₈N₂O) C, H, N.

(-)-(3a*S*)-1,8-Dibenzyl-3a-methyl-1,2,3,3a,8,8a-hexahydro-5-yl [(-)17]. Compound (-)16 (140 mg, 0.36 mmol) was dissolved in CH₂Cl₂ (2 mL), and boron tribromide (456 mg, 1.82 mmol) was added with stirring. The reaction mixture was stirred at room temperature for 1 h, MeOH (1 mL) was added dropwise, and the mixture was stirred for 10 min. After evaporation of solvent in vacuo, the residue was dissolved in H₂O (2 mL), basified with NaHCO₃, and extracted with Et₂O (3 \times 5 mL). The ether extract was dried over Na₂SO₄ overnight, evaporated, and chromatographed (CH₂Cl₂/MeOH = 10/1) to give (-)17 (120 mg, 90.0%) as a foam: [α]_D²⁰ -76.5° (*c* = 0.2, CHCl₃); ¹H NMR (CDCl₃) δ 7.40–7.00 (m, 10 H, Ar-H), 6.70–6.00 (m, 3H, C4-H, C6-H, C7-H), 4.40–3.90 (m, 2H, PhCH₂-N8), 3.80–3.60 (m, 2H, PhCH₂-N1), 2.65 (m, 2H, C2-H₂), 1.89 (m, 2H, C3-CH₂), 1.30 (s, 3H, C3a-CH₃); EI-MS *m/z* (relative intensity) 294 (MH⁺ - phenyl, 15), 380 (34), 265 (9.3), 250 (2.8), 91 (100). Anal. (C₃₃H₃₃N₃O₃·0.9 H₂O) C, H, N.

(-)-(3a*S*)-1,8-Dibenzyl-3a-methyl-1,2,3,3a,8,8a-hexahydro-5-yl *N*-Phenylcarbamate [(-)18]. Compound (-)17 (40.7 mg, 0.11 mmol) was dissolved in anhydrous ether (2 mL), and a piece of Na metal (approximately 1 mg) was added. The mixture was stirred at room temperature for 1 min, phenyl isocyanate (13.1 mg, 0.11 mmol) was added, and the mixture was stirred for 5 min. Evaporation of solvent gave a crude product which was directly chromatographed to give (-)18 (48 mg, 89.1%) as a gum: [α]_D²⁰ -59.1° (*c* = 0.7, CHCl₃); ¹H NMR (CDCl₃) δ 7.40–6.90 (m, 15H, Ar-H), 6.75 (d, *J* = 2.5 Hz, 1H, C4-H), 6.73 (d, *J* = 8.5 Hz, 1H, C6-H), 6.17 (d, *J* = 8.5 Hz, 1H, C7-H), 4.45 (s, 1H, C8a-H), 4.30–4.20 (AB, *J* = 16.6 Hz, 2H, PhCH₂-N8), 3.72 (s, 2H, PhCH₂-N1), 2.70 (m, 2H, C2-H₂), 1.90 (m, 2H, C3-H₂), 1.38 (s, 3H, C3a-CH₃); EI-MS *m/z* (relative intensity) 370 (MH⁺ - PhNHCO, 31), 354 (1.0), 279 (8.0), 264 (2.0), 91 (100). Anal. (C₂₇H₂₉N₃O₃) C, H, N.

(-)-(3a*S*)-1,8-Dibenzyl-3a-methyl-1,2,3,3a,8,8a-hexahydro-5-yl *N*-Methylcarbamate [(-)19]. Compound (-)16 (47.5 mg, 0.13 mmol) was dissolved in anhydrous ether (2 mL), and a piece of Na metal (approximately 1 mg) was added. The mixture was stirred at room temperature for 1 min, methyl isocyanate (14.6 mg, 0.26 mmol) was added, and the mixture stirred for 10 min. Evaporation of solvent gave a crude product which was directly chromatographed to give (-)19 (50.0 mg, 90.0%) as a gum: [α]_D²⁰ -58.2° (*c* = 0.7, CHCl₃); ¹H NMR (CDCl₃) δ 7.40–7.20 (m, 10H, Ar-H), 6.75 (d, *J* = 2.2 Hz, 1H, C4-H), 6.64 (d, *J* = 8.5 Hz, 1H, C6-H), 6.24 (d, *J* = 8.5 Hz, 1H, C7-H), 4.65 (s, 1H, N-H), 4.40 (s, 1H, C8a-H), 4.35–4.20 (AB, *J* = 16.6 Hz, 2H, PhCH₂-N8), 3.70 (s, 2H, PhCH₂-N1), 2.80 (d, *J* = 3.9 Hz, 3H, NHCH₃), 2.70 (m, 2H, C2-H₂), 1.90 (m, 2H, C3-H₂), 1.35 (s, 3H, C3a-CH₃); EI-MS *m/z* (relative intensity) 370 (MH⁺ - CH₃NHCO, 33), 354 (1.5), 279 (8.5), 264 (3.0), 91 (100). Anal. (C₂₇H₂₉N₃O₃) C, H, N.

(-)-(3a*S*)-3a-Methyl-1,2,3,3a,8,8a-hexahydro-5-yl *N*-Phenylcarbamate [(-)9]. Compound (-)18 (29.3 mg, 0.06 mmol) was dissolved in 2-propanol (1 mL), and Pd(OH)₂/C (5 mg) was added. The reaction mixture was

stirred under hydrogen at atmospheric pressure and room temperature for 60 h, and then the catalyst was filtered. Evaporation of solvent gave a residue which was chromatographed (CH₂Cl₂/MeOH = 10/1) to obtain the most polar component and this was crystallized from petroleum ether to provide compound (-)9 (12 mg, 64.7%) as crystals: mp 153–155 °C; [α]_D²⁰ -73.7° (*c* = 0.2, EtOH); ¹H NMR (CDCl₃/CD₃OD = 8/2) δ 7.50–7.00 (m, 5H, Ph-H), 6.90–6.50 (m, 3H, Ar-H), 4.80 (s, 1H, C8a-H), 3.10–2.75 (m, 2H, C2-H₂), 2.15–1.80 (m, 2H, C3-H₂), 1.45 (s, 3H, C3a-CH₃); EI-MS *m/z* (relative intensity) 232 (M⁺ - phenyl, 67), 217 (38), 190 (11), 174 (100), 160 (73); CI-MS (NH₃) *m/z* 310 (MH⁺). Anal. (C₂₇H₂₉N₃O₃) C, H, N.

(-)-(3a*S*)-3a-Methyl-1,2,3,3a,8,8a-hexahydro-5-yl *N*-Methylcarbamate [(-)10]. Compound (-)19 (34.0 mg, 0.08 mmol) was dissolved in 2-propanol (1 mL), and Pd(OH)₂/C (5 mg) was added. The reaction mixture was stirred under hydrogen at atmospheric pressure and room temperature for 48 h, and then the catalyst was filtered. Evaporation of solvent gave a residue which was chromatographed (CH₂Cl₂/MeOH = 10/1) to obtain the most polar component, compound (-)10 (12 mg, 60.7%), as a gum: [α]_D²⁰ -114.3° (*c* = 0.2, EtOH); ¹H NMR (CDCl₃/CD₃OD = 8/2) δ 6.90–6.55 (m, 3H, Ar-H), 5.18 (s, 1H, C8a-H), 2.95–2.70 (m, 2H, C2-H₂), 2.85 (d, *J* = 4.0 Hz, 3H, NCH₃), 2.10–1.80 (m, 2H, C3-H₂), 1.50 (s, 3H, C3a-CH₃); EI-MS *m/z* (relative intensity) 232 (M⁺ - methyl, 65), 217 (38), 189 (8.3), 174 (100); CI-MS (NH₃) *m/z* 248 (MH⁺). Anal. (C₂₇H₂₉N₃O₃) C, H, N.

(+)**9** and (+)**10** were synthesized from (+)**12** according to the same procedures used for making their antipodes. The ¹H NMR and MS of (+)**9** and (+)**10** and the intermediates (+)**14**, (+)**17**, (+)**18**, and (+)**19** were identical to those of their respective antipodes. The optical rotation values of the plus series compounds were equal and opposite to those of their minus enantiomers.

(-)-*N*⁸-Norphenserine (**7**) Made from Compound (-)18. Compound (-)18 (29.3 mg, 0.06 mmol) was dissolved in MeOH (1 mL), and Pd(OH)₂/C (5 mg) was added. The reaction mixture was stirred under hydrogen at atmospheric pressure and room temperature for 60 h, and then the catalyst was filtered. Evaporation of solvent gave a residue which was chromatographed (CH₂Cl₂/MeOH = 10/1) to obtain the most polar component and this was crystallized from petroleum ether to provide compound **7** (13.6 mg, 70.0%) as crystals: The mp, optical rotation, ¹H NMR, and MS were identical with those reported in the literature.⁸

(-)-(3*S*)-3-Methyl-3-[2'-(amino)ethyl]-2,3-dihydro-1*H*-indol-5-yl *N*-Phenylcarbamate (**20**). Compound (-)18 (58.6 mg, 0.12 mmol) was dissolved in a mixture of MeOH (2 mL), H₂O (2 mL), and TFA (1 mL), and Pd(OH)₂/C (10 mg) was added. The reaction mixture then was stirred under hydrogen at atmospheric pressure and room temperature for 1.5 h, and thereafter the catalyst was filtered. The filtrate was evaporated in vacuo to give a residue which was dissolved in H₂O, basified by Na₂CO₃, extracted with CH₂Cl₂, and then dried over Na₂SO₄. After removal of solvent, the residue was chromatographed on preparative TLC (silica gel) (CH₂Cl₂/MeOH = 10/1) to give the most polar and also major component **20** (7.4 mg, 20%) as a syrup: [α]_D²⁰ -63.2° (*c* = 0.1, CHCl₃); ¹H NMR (CDCl₃/CD₃OD = 8/2) δ 7.40–7.00 (m, 5H, Ph-H), 6.85–6.70 (m, 2H, C4-H, C6-H), 6.50 (d, *J* = 8.5 Hz, 1H, C7-H), 3.75 and 3.65 (AB, *J* = 12 Hz, 2H, C2-H₂), 3.40–3.20 (AB, *J* = 12 Hz, CH₂N), 1.65 (m, 2H, C3-CH₂), 1.30 (s, 3H, C3-CH₃); CI-MS (NH₃) *m/z* 312 (MH⁺). Anal. (C₁₃H₁₆N₂O₃) C, H, N.

Quantitation of Anticholinesterase Activity. The action of compounds **3**, **4**, and (-) and (+)**9**, **10**, **18**, and **19** to inhibit the ability of freshly prepared human AChE and BChE, derived from plasma and whole red blood cells, respectively, to enzymatically degrade the specific substrates acetyl(β -methyl)thiocholine and *s*-butyrylthiocholine (0.5 mmol/L) (Sigma Chemical Co., St. Louis, MO) were quantified. Compounds were dissolved in Tween 80/EtOH, 3:1 (v:v; <150 μ L total volume), and were diluted in 0.1 M Na₃PO₄ buffer (pH 8.0) in half-log concentrations to provide a final range spanning 0.3

nM to 30 mM. Tween 80/EtOH was diluted in excess of 1 in 5000 and had no inhibitory action on either AChE or BChE.

Freshly collected blood was centrifuged (10000g, 10 min, 4 °C), and plasma was removed and diluted 1:125 with 0.1 M Na₃PO₄ buffer (pH 7.4). Whole red blood cells were washed five times in isotonic saline, lysed in 9 volumes of 0.1 M Na₃PO₄ buffer (pH 7.4) containing 0.5% Triton-X (Sigma), and then diluted with an additional 19 volumes of buffer to a final dilution of 1:200. Analysis of anticholinesterase activity, utilizing a 25 μL sample of each enzyme preparation, was undertaken at their optimal working pH, 8.0, in 0.1 M Na₃PO₄ buffer (0.75 mL total volume). Compounds were preincubated with enzymes (30 min, room temperature) and then were incubated with their respective substrates and 5,5'-dithiobis-2-nitrobenzoic acid (25 min, 37 °C). Production of a yellow thionitrobenzoate anion was measured by spectrophotometer at 412 nm λ. To correct for nonspecific substrate hydrolysis, aliquots were co-incubated under conditions of absolute enzyme inhibition (by the addition of 1 × 10⁻⁵ M physostigmine [(-)-2]), and the associated alteration in absorbance subtracted from that observed through the concentration range of each test compound. Each was analyzed on four separate occasions and assayed alongside (-)-phenserine [(-)-1] and (-)-physostigmine [(-)-2] as control and external standards whose activity we have previously reported.^{1,2,7,8,18,21}

The enzyme activity at each concentration of test compound was expressed as a percent of activity in the absence of compound, transformed into a logit format (logit = %activity/(100 - %activity)) and then plotted as a function of its log concentration. Inhibitory activity was calculated as an IC₅₀, defined as the concentration of compound (nM) required to inhibit 50% of enzymatic activity, and determined from a correlation between log concentration and logit activity.

Acknowledgment. The authors are extremely grateful to Noel F. Whittaker, Laboratory of Bioorganic Chemistry, National Institute of Diabetes and Digestive and Kidney Diseases, NIH, for undertaking the MS analysis of the compounds reported herein and are indebted to Dr. Amy Newman, Medicinal Chemistry, National Institute on Drug Abuse, NIH, for use of NMR and optical rotation equipment.

References

- Greig, N. H.; Pei, X. F.; Soncrant, T. T.; Ingram, D. K.; Brossi, A. Phenserine and Ring C Hetero-Analogues: Drug Candidates for Treatment of Alzheimer's Disease. *Med. Res. Rev.* **1995**, *15*, 3-31.
- Brossi, A.; Pei, X. F.; Greig, N. H. Invited Review. Phenserine, a Novel Anticholinesterase Related to Physostigmine: Total Synthesis and Biological Properties. *Aust. J. Chem.* **1996**, *49*, 171-181.
- Greig, N. H.; Ingram, D.; Wallace, W. C.; Utsuki, T.; Yu, Q. S.; Holloway, H. W.; Pei, X. F.; Haroutunian, V.; Lahiri, D.; Brossi, A.; Soncrant, T. T. Phenserine: a selective, long-acting and brain-directed acetylcholinesterase inhibitor affecting cognition and β-APP processing. In *Alzheimer's Disease: Molecular Biology to Therapy*; Becker, B., Giacobini, E., Robert, P., Eds.; Birkhäuser, Boston, 1996; pp 231-237.
- Haroutunian, V.; Greig, N. H.; Utsuki, T.; Davis, K. L.; Wallace, W. C. Pharmacological modulation of Alzheimer's β-amyloid precursor protein levels in the CSF of rats with forebrain cholinergic system lesions. *Mol. Brain Res.* **1997**, *46*, 161-168.
- McMahon, B. E. *A. Burger's Medicinal Chemistry Part 1*; Wiley-Interscience: New York, **1970**, 50.
- Testa, B.; Jenner, P. *Drug Metabolism: Chemical and Biochemical Aspects*; Marcel Dekker Inc.: New York, 1976; Vol. 4.
- Yu, Q. S.; Atack, J. R.; Rapoport, S. I.; Brossi, A. Synthesis and Anticholinesterase Activity of (-)-N¹-Norphysostigmine, (-)-Eseramine, and Other N¹-Substituted Analogues of (-)-Physostigmine. *J. Med. Chem.* **1988**, *31*, 2297-2300.
- Yu, Q. S.; Pei, X. F.; Holloway, H. W.; Greig, N. H.; Brossi, A. Total Syntheses and Anticholinesterase Activities of (3aS)-N⁸-Norphysostigmine, (3aS)-N⁸-Norphenserine, Their Antipodal Isomers, and Other N⁸-Substituted Analogues. *J. Med. Chem.* **1997**, *40*, 2895-2901.
- Pei, X. F.; Greig, N. H.; Bi, S.; Brossi, A.; Toome, V. Inhibition of Human Acetylcholinesterase by Unnatural (+)-(3aR)-N¹-Norphysostigmine and Arylcarbamate Analogues. *Med. Chem. Res.* **1995**, *5*, 265-270.
- Yu, Q. S.; Schönenberger, B.; Brossi, A. Reactions of (-)-Physostigmine and (-)-N-Methylphysostigmine in Refluxing Butanol and at High Temperature: Facile Preparation of (-)-Eseroline. *Heterocycles* **1987**, *26*, 1271-1275.
- Auterhoff, H.; Hamacher, H. Die Farbreaktion des Eserines. *Arch. Pharm.* **1967**, *300*, 849-856.
- Hemsworth, B. A.; West, G. B. Anticholinesterase Activating of Some Degradation Products of Physostigmine. *J. Pharm. Sci.* **1970**, *59*, 118-120.
- Schönenberger, B.; Jacobson, A. E.; Brossi, A.; Streaty, R.; Klee, W. A.; Flippen-Anderson, J. L.; Gilardi, R. Comparison of (-)-Eseroline with (+)-Eseroline and Dihydroseco Analogues in Antinociceptive Assays: Confirmation of Rubreserine Structure by X-ray Analysis. *J. Med. Chem.* **1986**, *29*, 2268-2273.
- Poobrasert, O.; Chai, H.; Pezzuto, J. M.; Cordell, G. A. Cytotoxic Degradation Product of Physostigmine. *J. Nat. Prod.* **1996**, *59*, 1087-1089.
- Hoshino, T.; Tamura, K. Synthesizche Versuche in der Indol-Gruppe. X. Über die Synthese des Eserin-Ringsystems. *Liebigs Ann. Chem.* **1932**, *500*, 42-52.
- Pei, X. F.; Greig, N. H.; Flippen-Anderson, J. L.; Bi, S.; Brossi, A. Total Synthesis of Racemic and Optically Active Compounds Related to Physostigmine and Ring-C Heteroanalogues from 3-[2'-(Dimethylamino)ethyl]-2,3-dihydro-5-methoxy-1,3-dimethyl-1H-indol-2-ol. *Helv. Chim. Acta* **1994**, *77*, 1412-1421.
- Baiker, A.; Richarz, W. Synthesis of Long Chain Aliphatic Amines From The Corresponding Alcohols. *Tetrahedron Lett.* **1977**, *22*, 1937-1938.
- He, S.; Brossi, A. N-Demethylation of (±)-6β-Acetoxy-3-Tropinone Synthesis of (±)-6β-Acetoxyntropine. *J. Heterocycl. Chem.* **1991**, *28*, 1741-1745.
- Ijima, S.; Greig, N. H.; Garofalo, P.; Spangler, E. L.; Brossi, A.; Ingram, D. K. Phenserine: a Physostigmine Derivative that is a Long-Acting Inhibitor of Cholinesterase and Demonstrates a Wide Dose Range for Attenuating a Scopolamine-Induced Learning Impairment of Rats in a 14-unit T-Maze. *Psychopharmacology* **1993**, *112*, 415-420.
- Marta, M.; Castellano, C.; Oliverio, A.; Pavone, F.; Pagella, P. G.; Brufani, M.; Pomponi, M. New Analogues of Physostigmine: Alternative Drugs for Alzheimer's Disease? *Life Sci.* **1988**, *43*, 1921-1928.
- Yu, Q. S.; Greig, N. H.; Holloway, H. W.; Utsuki, T.; Pei, X. F.; Brossi, A. (-)-(3aS)-Eseroline Carbamate, a Close Analogue of Physostigmine. *Med. Chem. Res.* **1997**, *7*, 116-122.
- Marta, M.; Gatta, F.; Pomponi, M. Physostigmine Analogues Anticholinesterases: Effects of the Lengthening of the N-Carbamic Chain on the Inhibition Kinetics. *Biochim. Biophys. Acta* **1992**, *1120*, 262-266.
- Somani, S. M.; Khalique, A. Pharmacokinetics and Pharmacodynamics of Physostigmine in the Rat after Intravenous Administration. *Drug Metab. Dis.* **1987**, *15*, 627-633.
- Becker, R.; Moriearty, P.; Unni, L. The Second Generation of Cholinesterase Inhibitors: Clinical and Pharmacological Effects. In *Cholinergic Basis of Alzheimer's Disease*; Becker, R., Giacobini, E., Eds.; Birkhäuser: Boston, 1991; pp 263-296.
- Yu, Q. S.; Liu, C.; Brzostowska, M.; Chrisey, L.; Brossi, A.; Greig, N. H.; Atack, J. R.; Soncrant, T. T.; Rapoport, S. I.; Radunz, H. E. Physovenines: Efficient Synthesis of (-) and (+)-Physovenine and Synthesis of Carbamate Analogues of (-)-Physovenine. Anticholinesterase Activity and Analgesic Properties of Optically Active Physovenines. *Helv. Chim. Acta* **1991**, *74*, 761-766.
- He, X. S.; Greig, N. H.; Rapoport, S. I.; Brossi, A.; Li, Y. Q.; Yu, Q. S. Thiaphysovenine and Carbamate Analogues: A New Class of Potent Inhibitors of Cholinesterases. *Med. Chem. Res.* **1992**, *2*, 229-237.
- Brzostowska, M.; He, X. S.; Greig, N. H.; Brossi, A. Selective inhibition of acetyl- and butyrylcholinesterases by phenylcarbamates of (-)-eseroline, (-)-(N1)-noreseroline and physovenol. *Med. Chem. Res.* **1992**, *2*, 238-246.
- Pei, X. F.; Greig, N. H.; Brossi, A. Preparation and Selective Inhibition of Human Butyrylcholinesterase by N1-Phenylethyl-norphysostigmine analogues. *Med. Chem. Res.* **1995**, *5*, 455-461.
- Soreq, H.; Zakut, H. *Human Cholinesterases and Anticholinesterases*; Academic Press: New York, 1993.
- Chatonnet, A.; Lockridge, O. Comparison of Butyrylcholinesterase and Acetylcholinesterase. *Biochem. J.* **1989**, *260*, 625-634.
- Massoulie, J.; Pezzementi, L.; Krejci, E.; Vellette, F. M. Molecular and Cellular Biology of Cholinesterase. *Prog. Neurobiol.* **1993**, *41*, 31-91.
- Ben-Aziz, R.; Gnatt, A.; Prody, C. A.; Lev-Lehman, E.; Neville, L.; Seidman, S.; Ginzberg, D.; Soreq, H.; Lapidot-Lifson, Y.; Zakut, H. Differential Codon Usage and Distinct Surface Prob-

- abilities in Human Acetylcholinesterase and Butyrylcholinesterase. In *Cholinesterases: Structure, Function, Mechanisms, Genetics and Cell Biology*; Massoulie, J., Bacou, F., Barnard, A., Chattonet, A., Doctor, B. P., Quinn, D. M., Eds.; ACS Books: Washington, DC, 1991; pp 172–178.
- (33) Giacobini, E. Cholinesterase Inhibitors do more than Inhibit Cholinesterase. In *Alzheimer's Disease: Molecular Biology to Therapy*; Becker, R., Giacobini, E., Eds.; Birkhäuser: Boston, 1996; pp 188–204.
- (34) Mesulam, M. M. Butyrylcholinesterase in Alzheimer's Disease. In *Alzheimer's Disease: Therapeutic Strategies*; Becker, R., Giacobini, E., Eds.; Birkhäuser: Boston, 1994; pp 79–83.
- (35) Barber, K. L.; Mesulum, M. M.; Krafft, G. A.; Klein, W. L. Butyrylcholinesterase Alters the Aggregation State of β -Amyloid. *Proc. Soc. Neurosci.* **1996**, *26*, 1172.

JM9800494